

Unsupervised Unstained Cell Detection using SIFT Keypoint Clustering and Laplacian Boundary Potential (2)

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Outlines

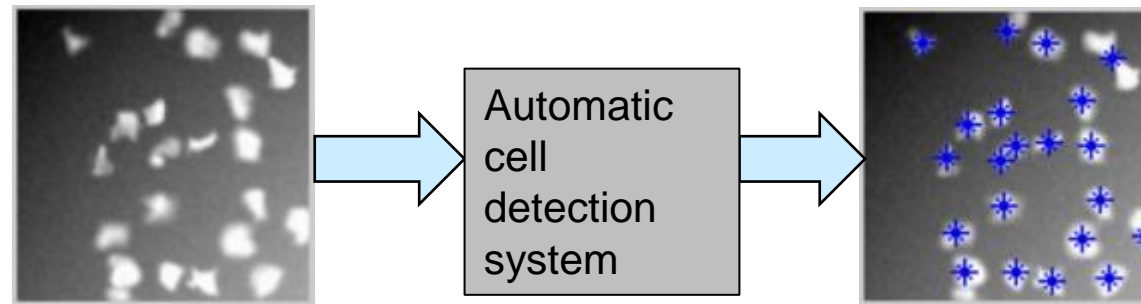
- Reminder of our supervised cell detection system
- Unsupervised approach
- Evaluation
- Conclusion & outlook

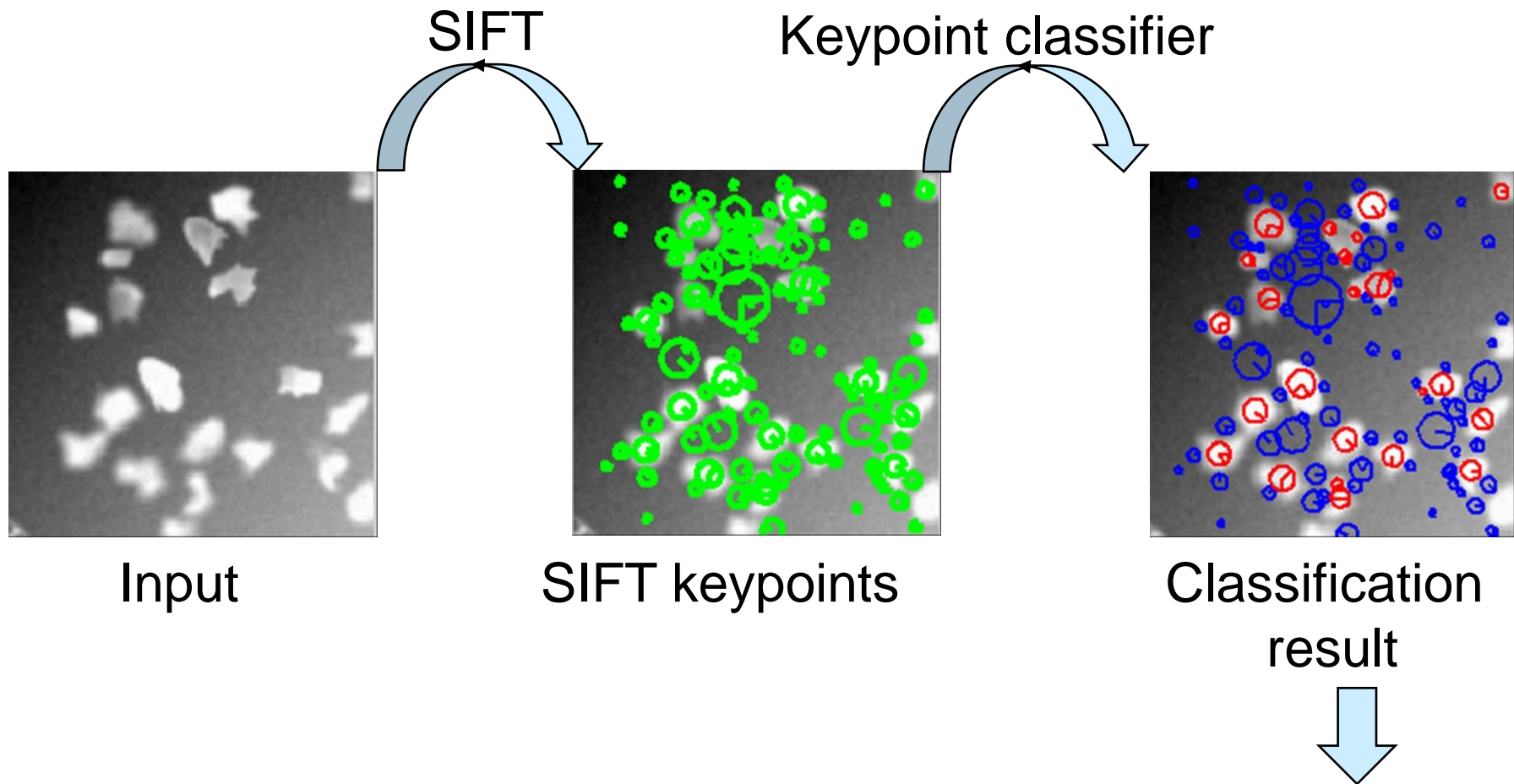
Reminder

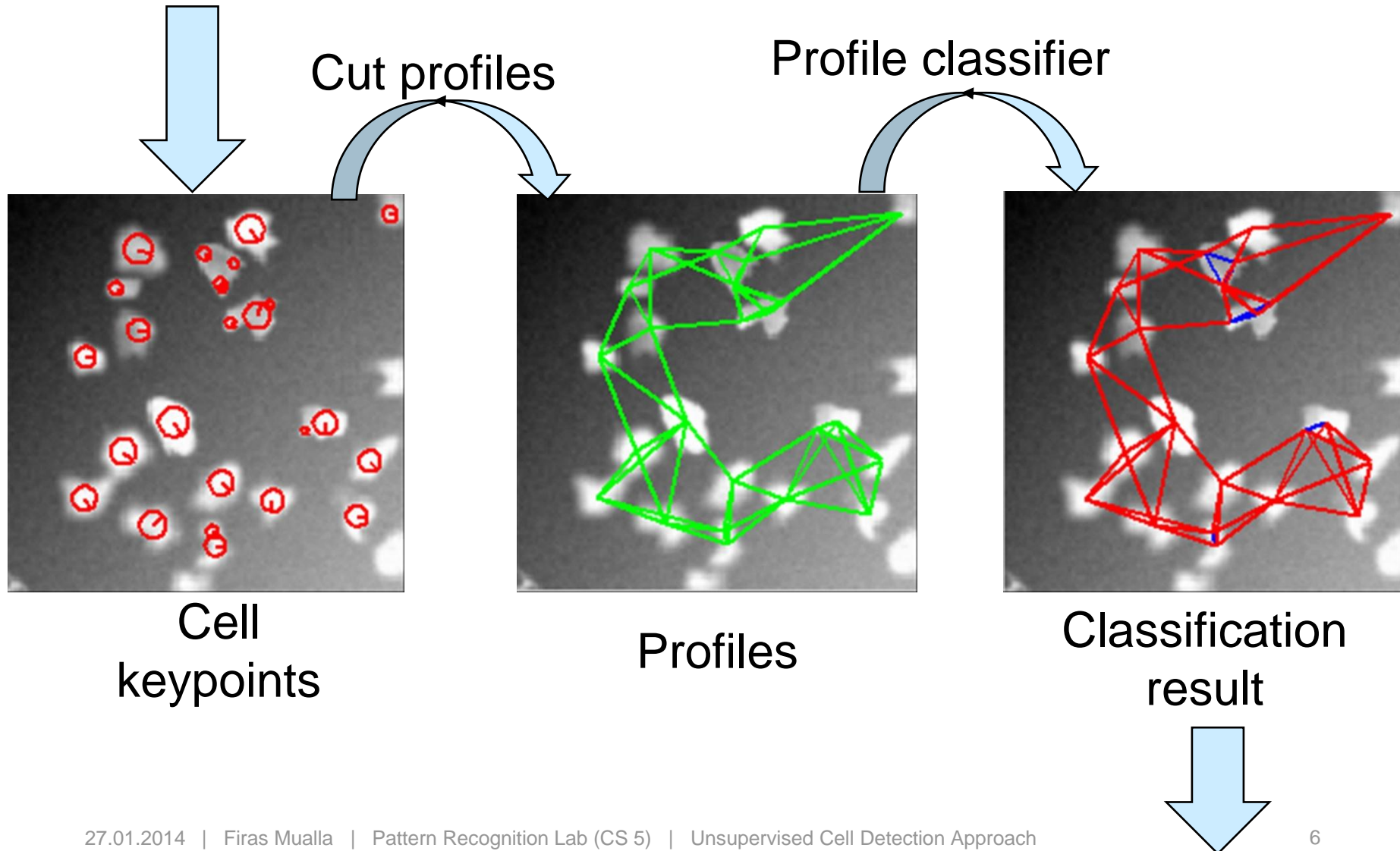


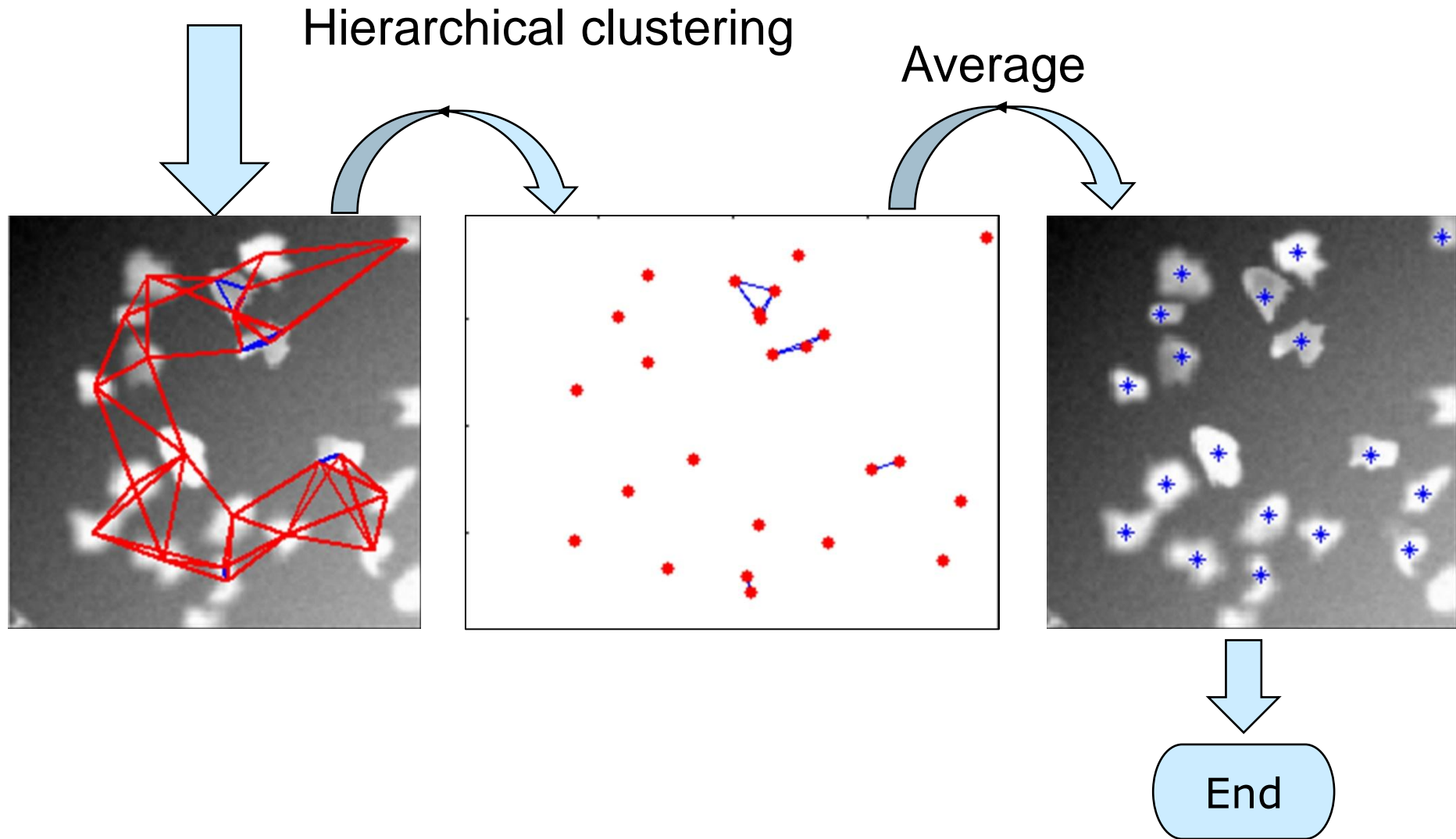


The system











So far!

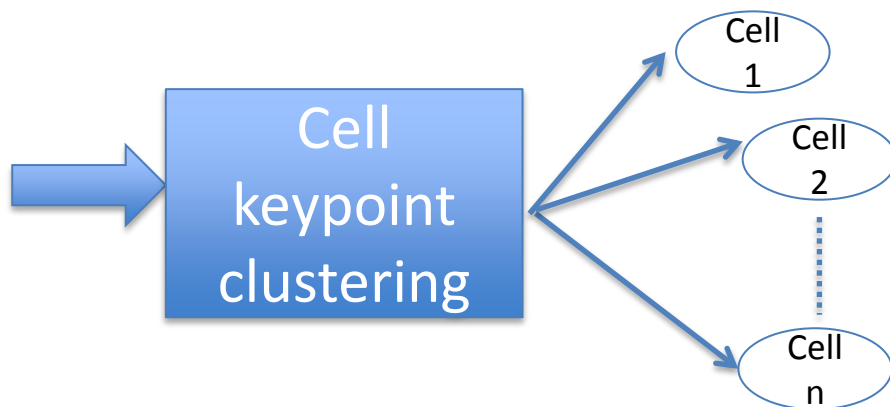
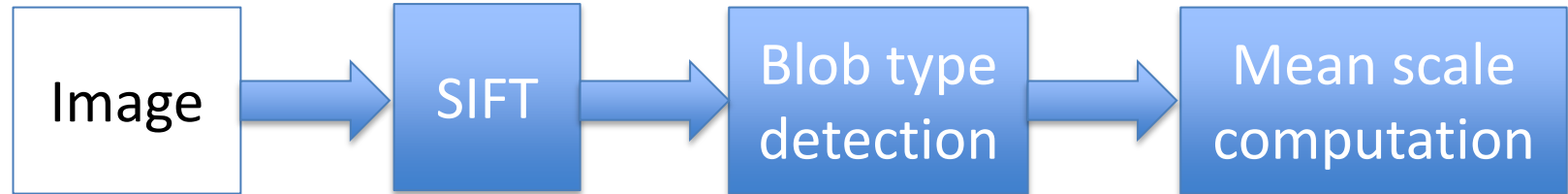
- + Fully automatic cell detection
- + High detection rate
- + High scores in scale-, orientation-, and illumination-invariance
- + General design
- Needs training



Unsupervised approach



System overview



Blob type detection



Positive

or



Negative

?

$$\omega_i = \frac{s(\mathbf{p}_i)}{\text{score}(\mathbf{p}_i)}$$

- $s(\mathbf{p}_i)$: scale of the keypoint \mathbf{p}_i .
- $\text{score}(\mathbf{p}_i)$: non-circularity measure.

> 0.5

< 0.5

$$\text{PositiveFit} = \frac{\sum_{i=1}^N \omega_i |DOG(\mathbf{p}_i)| H(DOG(\mathbf{p}_i))}{\sum_{i=1}^N \omega_i |DOG(\mathbf{p}_i)|}$$

- $\mathbf{p}_i, i = 1..N$ are the keypoints in the image.
- $DOG(\mathbf{p}_i)$: difference of Gaussians value.
- H is the Heaviside step function.

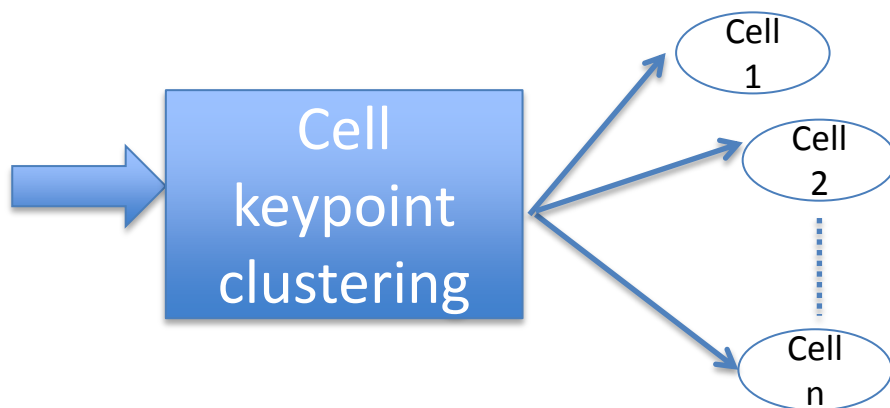
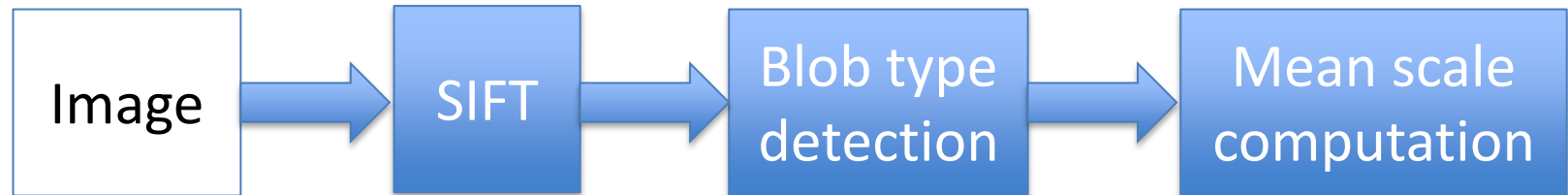


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System overview



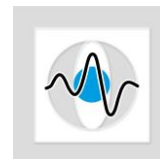
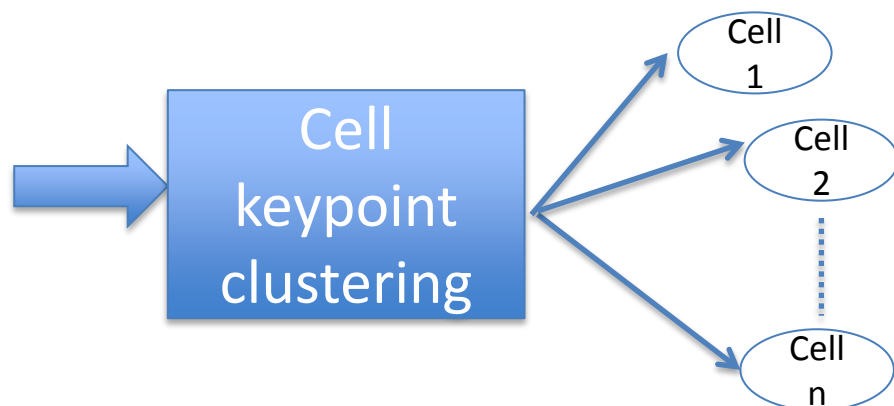
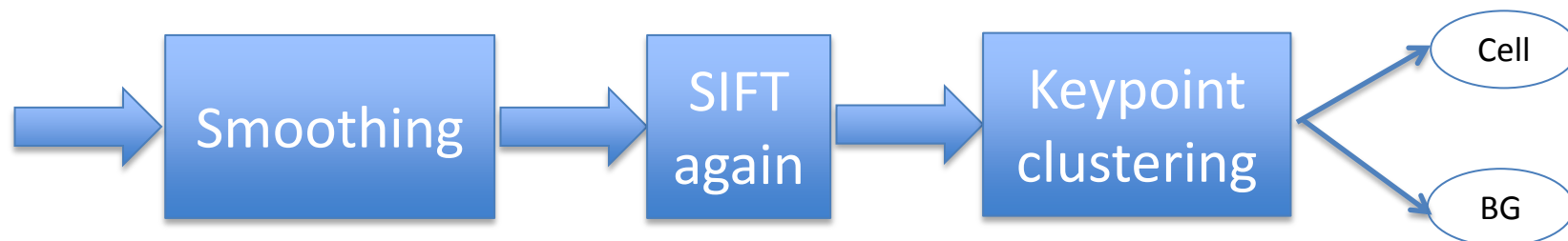
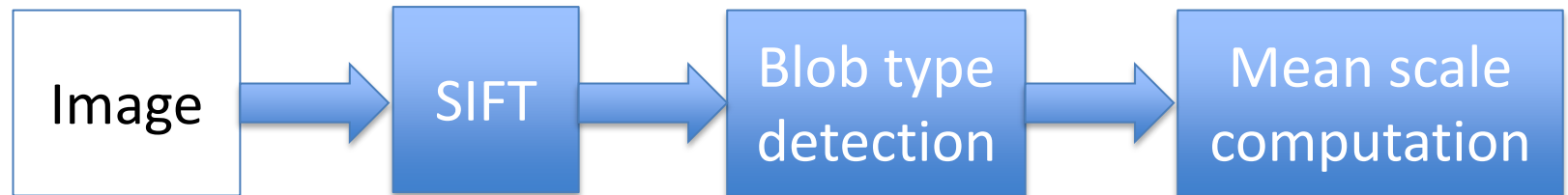
Mean scale computation

$$SE = \frac{\sum_{i=1}^M |DOG(\mathbf{p}_i)| s(\mathbf{p}_i)}{\sum_{i=1}^M |DOG(\mathbf{p}_i)|}$$

- $s(\mathbf{p}_i)$: scale of the keypoint \mathbf{p}_i .
- $DOG(\mathbf{p}_i)$: difference of Gaussians value.
- $\mathbf{p}_i, i = 1..M$ are the one-sided keypoints (after blob type detection).



System overview

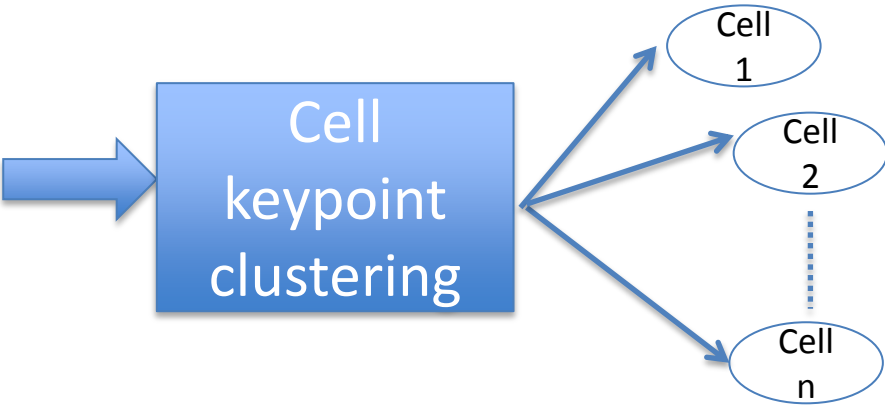
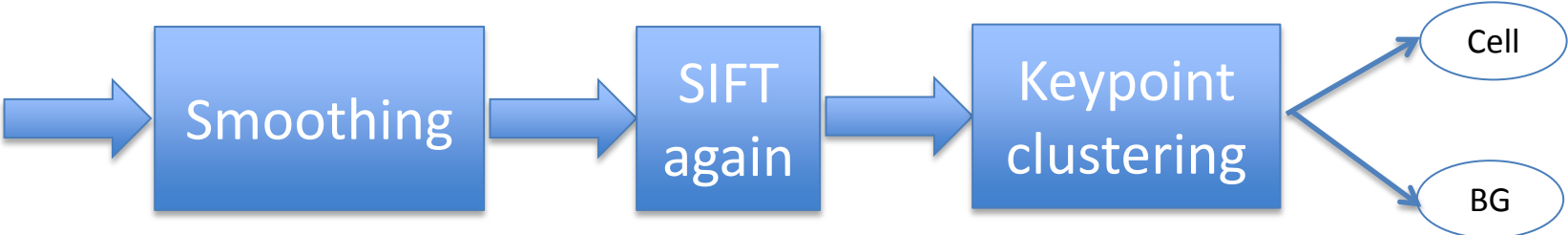
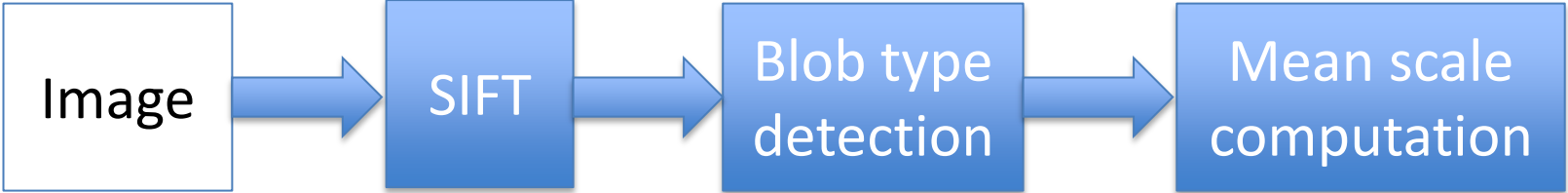


Smoothing

- Image is smoothed with a Gaussian kernel.
- Its standard deviation is the previous mean scale.
- Applying SIFT again:
 - The new keypoints are more stable.
 - Their number is considerably less.



System overview

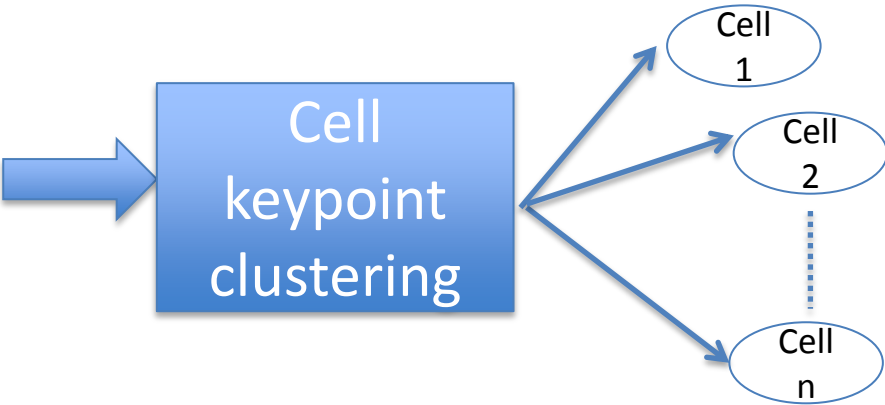
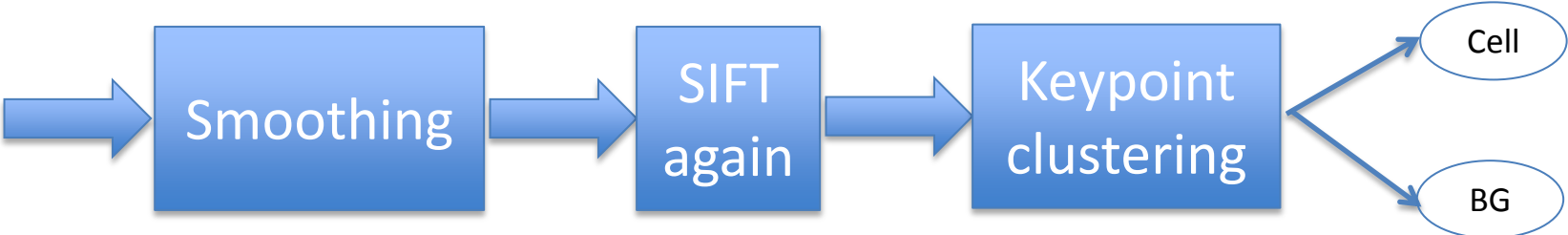
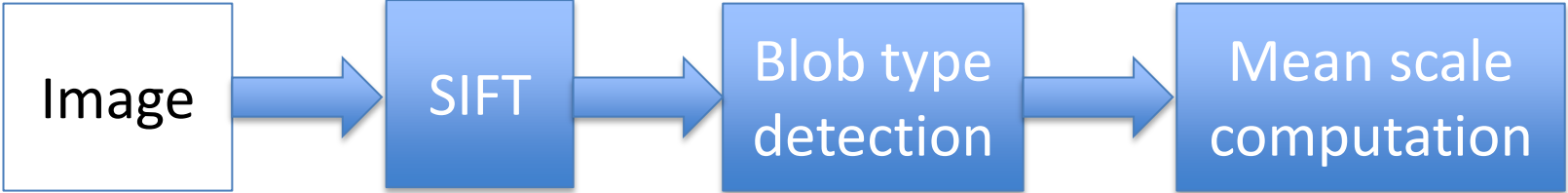




Keypoint clustering

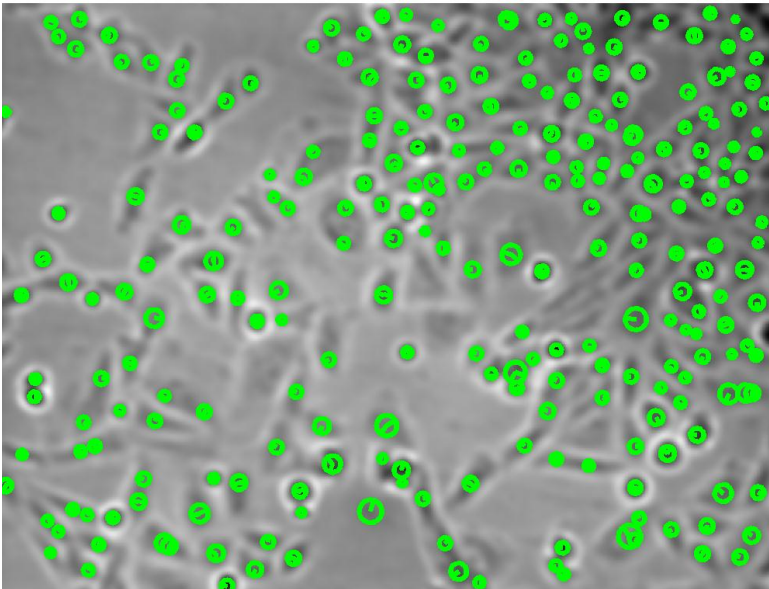
- K-means clustering
- Clusters which correspond to 1D Otsu thresholding of the DOG values are used as initialization.
- City-block distance measure
- Modality-specific features:
 - DOG and score were used in phase contrast.
 - DOG and intensity were used in bright-field.
- Applying SIFT again seems to have considerable effect on the result!

System overview

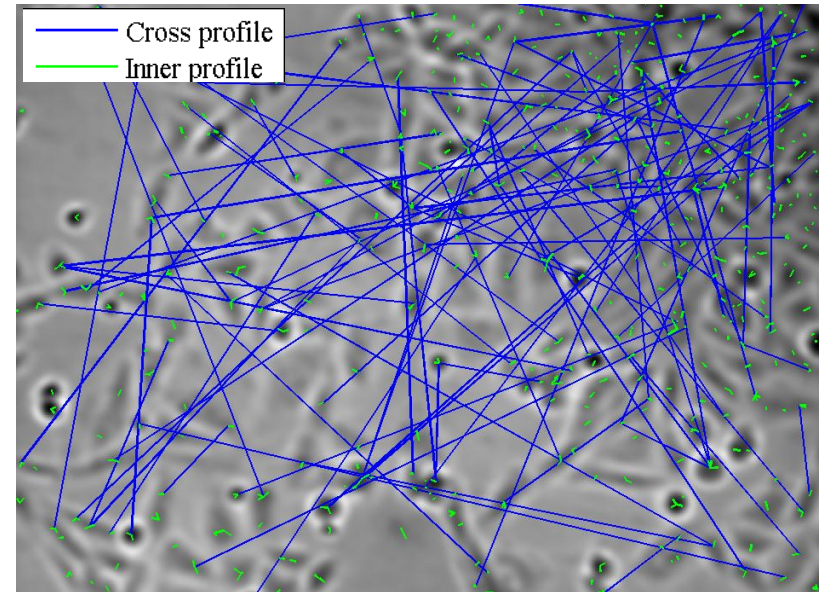


Cell keypoint clustering

- Back to **intuition**:



Cell SIFT keypoints



Artificial inner and cross profiles



Sample inner profiles

- Randomly sample M_1 keypoints from the resulting cell keypoints.
 - Use a triangular distribution in scale.
 - M_1 was set to 100 in our experiments.
- For each keypoint \mathbf{p}_i , pick a random angle θ_i and extract a profile between the following two points:
 - \mathbf{p}_i
 - $\mathbf{p}_i + (s(\mathbf{p}_i)\cos(\theta_i), s(\mathbf{p}_i)\sin(\theta_i))$
- Assign label "inner" to the extracted profiles.



Sample cross profiles

- Compute the pairwise Euclidean distance matrix of the cell keypoints.
- Randomly pick M_2 distances more than $C \cdot SE$:
 - M_2 was set to 100.
 - C was set to 10.
- Assign label "cross" to the corresponding profiles.





Boundary potential

- Laplacian of Gaussian L of the **smoothed** image is used as boundary potential.
 - $L(\text{blob-type} * L > 0) = 0$, (blob-type = +1 or -1)
 - $L = |L|$

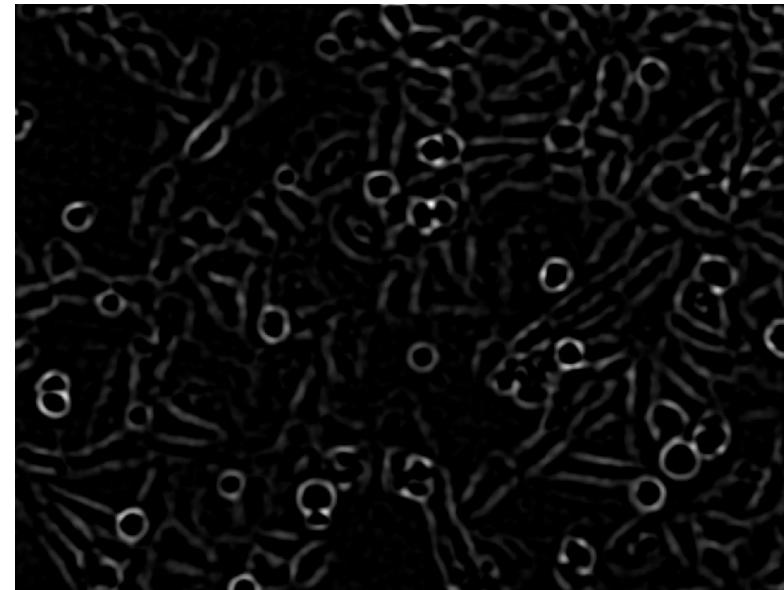
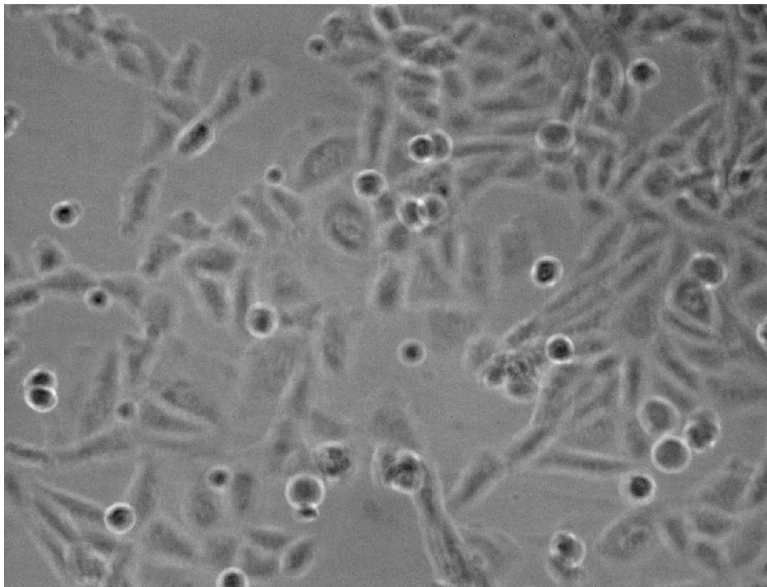


Image of CHO cells and its boundary potential



Training

- Extract features which are invariant to profile length.
 - Currently we use $\max(L)$ along the extracted profile.
- Use the extracted features to train a Bayesian classifier with Gaussian class conditional densities.



Cell keypoint clustering

- For each cell keypoint:
 - Find its K-nearest neighbors ($K=3$).
 - Classify the corresponding profiles using the Bayesian classifier.
- Similar to the supervised approach:
 - Agglomerative hierarchical clustering
 - Linkage method: average

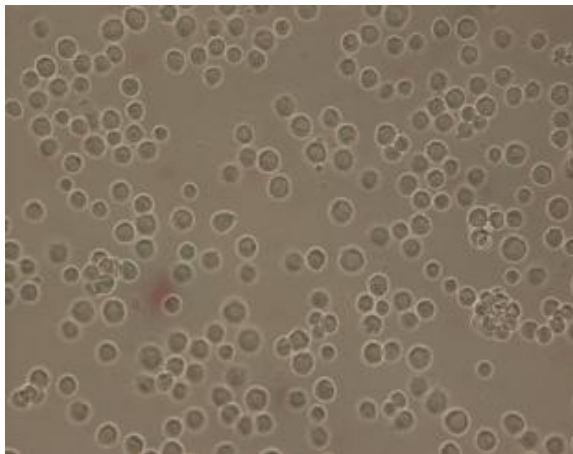


Evaluation

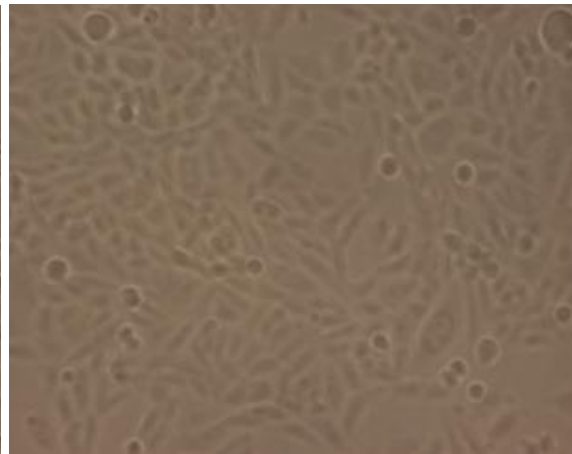


Bright-field

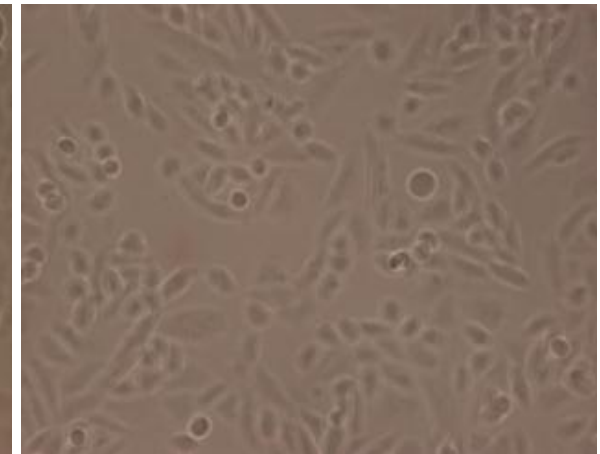
- 16 images
- Three cell lines
- More than 3500 manually labeled cells
- Ground truth type: border delineation



Sf21 cells



CHO cells

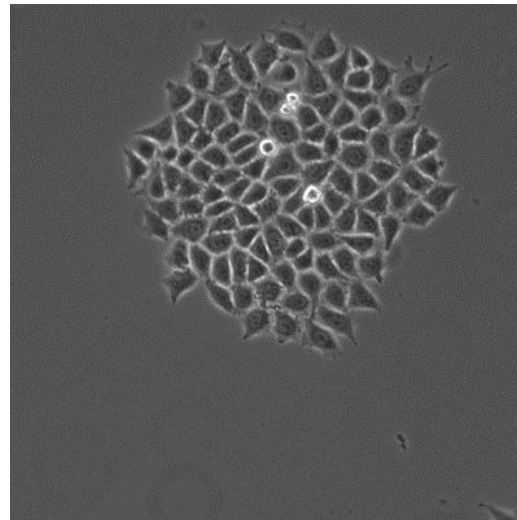


L929 cells



Phase contrast

- 11 images
- One cell line
- More than 1100 cells
- Ground truth type: dot at each cell center



HeLa cells



Results

	CHO	L929	Sf21	HeLa
F-Measure	80.58 %	84.31 %	88.26 %	85.72 %



Comparison with state-of-the-art: bright-field

	Mualla et al.[1]	Our approach	Becattini et al.[2]
F-measure	87.85 %	81.24 %	69.64 %

One CHO image was used to train [1] and the rest of the CHO images were used for evaluating the three approaches

[1] Mualla F, Schöll S, Sommerfeldt B, Maier A, Hornegger J (2013) Automatic cell detection in bright-field microscope images using SIFT, random forests, and hierarchical clustering. *IEEE Trans Med Image* 32(12):2274–2286.

[2] Becattini et al “A novel framework for automated targeting of unstained living cells in bright field microscopy,” in *Proceedings of the IEEE International Symposium on Biomedical Imaging: From Nano to Macro*, April 2011, pp. 195–198.



Comparison with state-of-the-art: phase contrast

	Arteta et al.[3]	Our approach
F-Measure	87.95 %	85.74 %

[3] C. Arteta, V.S. Lempitsky, J.A. Noble, and A. Zisserman, "Learning to detect cells using non-overlapping extremal regions", ;in Proc. MICCAI (1), 2012, pp.348-356.



Conclusions & outlook



Conclusions

- We have an unsupervised approach which has competitive performance compared to supervised state-of-the-art approaches on bright-field and phase contrast microscopy.
- Smoothing with mean image scale seems to provide:
 - Stable keypoints
 - Very good boundary potential between cells
- Use extreme cases when you do not have ground truth.



Outlook

- More features?
- More clever inner/cross profile selection?
- Bottleneck: Background / cell clustering



Thank you very much!